

### PTCL: morphology and pathobiology Anaplastic large cell lymphoma



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#### MATURE T-CELL AND NK-CELL NEOPLASMS

T-cell prolymphocytic leukaemia	9834/3
T-cell large granular lymphocytic leukaemia	9831/3
Chronic lymphoproliferative disorder of NK-cells	9831/3
Aggressive NK cell leukaemia	9948/3
Systemic EBV positive T-cell lymphoproliferati	ve
disease of childhood	9724/3
Hydroa vaccineforme-like lymphoma	9725/3
Adult T-cell leukaemia/lymphoma	9827/3
Extranodal NK/T cell lymphoma, nasal type	9719/3
Enteropathy-associated T-cell lymphoma	9717/3
Hepatosplenic T-cell lymphoma	9716/3
Subcutaneous panniculitis-like	
T-cell lymphoma	9708/3
Mycosis fungoides	9700/3
Sézary syndrome	9701/3

Primary cutaneous CD30 positive T-cell lymphoproliferative disorders	
Lymphomatoid papulosis	9718/1
Primary cutaneous anaplastic large cell lymphoma	9718/3
Primary cutaneous gamma-delta T-cell lymphoma	9726/3
Primary cutaneous CD8 positive aggressive epidermotropic cytotoxic T-cell lymphoma	9709/3
Primary cutaneous CD4 positive small/medium T-cell lymphoma	9709/3
Peripheral T-cell lymphoma, NOS	9702/3
Angioimmunoblastic T-cell lymphoma	9705/3
Systemic	



<sup>(\*)</sup> Feldman et a., Mod Pathol 2010, 23:593602.

# Morphologic spectrum of "ALK+ ALCL







nucleophosmin.

#### Translocations and fusion proteins involving the ALK gene in ALK+ ALCL

#### Anaplastic Large-Cell Lymphoma Inghirami and Pileri

Table 1      Chromosomal translocations involving the ALK gene in human lymphoma						
Disease	Chromosomal abnormalities	Fusion protein (kDa)	Partner gene	Frequency (%)	ALK IHC stains	Principal references
ALCL-DLBCL	t(2;5)(p23;q35)	NPM-ALK (80)	NPM1	75-80	Cyto/nuclear Nuclear	7, 18
ALCL-IMT	t(1;2)(q25;p23)	TPM3-ALK (104) (104)	ТРМЗ	12-18	Cyto	45
ALCL	t(2;3)(p23;q21)	TGF-ALK 113,97.85)	TFG	2	Cyto	46, 47
ALCL-IMT	inv(2)(p23;q35)	ATIC-ALK (96)	ATIC	2	Cyto	48, 73
ALCL-IMT-DLBCL	t(2;17)(p23;q23)	CLTC1-ALK (250)	CLTL1	2	Cyto	49
ALCL	t(2;17)(p23;q25)	AL017-ALK (ND)	AL017	<1	Cyto	51
ALCL	t(2;X)(p32;q11-12)	MSN-ALK (125)	MSN	<1	Cyto	50, 73
ALCL-IMT	t(2;19)(p23;p13)	TPM4-ALK(95-105)	TPM4	<1	Cyto	73
ALCL	t(2;22)(p23;q11.2)	MYH9-ALK (220)	МҮН9	<1	Cyto	52

Abbreviations: ALCL, anaplastic large-cell lymphoma; ALK, anaplastic lymphoma kinase; ATIC, 5-aminoimidazole-4-carboxamide ribonucleotide formyltransferase/IMP cyclohydrolase; CLTL1, Clathrin heavy chainlike1; cyto, cytoplasmic; DLBCL, diffuse large B-cell lymphoma; MSN, moesin; NPM,



195





Anaplastic large-cell lymphoma. Inghirami G and Pileri SA. Sem Diagn Pathol 2011; 28:190-201. ALK Kinase Domain Mutations in Primary Anaplastic Large Cell Lymphoma: Consequences on NPM-ALK Activity and Sensitivity to Tyrosine Kinase Inhibitors

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### ALK<sup>+</sup> ALCL

#### Anaplastic large-cell lymphoma

Seminars in Diagnostic Pathology (2011) 28, 190-201

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Small Molecule ALK-Tyrosine Kinase Inhibitors



### medicine

### PDGFR blockade is a rational and effective therapy for NPM-ALK-driven lymphomas

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## ALCL: ALK-positive and ALK-negative status

#### ALK-positive: most frequent in the first three decades of life<sup>1</sup>



#### Overall survival of systemic ALK according to ALK status<sup>2</sup>





### A novel patient-derived tumorgraft model with TRAF1-ALK anaplastic large-cell lymphoma translocation Leukemia (2015), 1–12

F Abate<sup>1,2,3,21</sup>, M Todaro<sup>3,21</sup>, J-A van der Krogt<sup>4,21</sup>, M Boi<sup>3,5</sup>, I Landra<sup>3</sup>, R Machiorlatti<sup>3</sup>, F Tabbò<sup>3</sup>, K Messana<sup>3</sup>, C Abele<sup>3</sup>, A Barreca<sup>3</sup>, D Novero<sup>3</sup>, M Gaudiano<sup>3</sup>, S Aliberti<sup>3</sup>, F Di Giacomo<sup>3</sup>, T Tousseyn<sup>6</sup>, E Lasorsa<sup>3</sup>, R Crescenzo<sup>3</sup>, L Bessone<sup>3</sup>, E Ficarra<sup>1</sup>, A Acquaviva<sup>1</sup>, A Rinaldi<sup>5</sup>, M Ponzoni<sup>7</sup>, DL Longo<sup>8</sup>, S Aime<sup>8</sup>, M Cheng<sup>9,22</sup>, B Ruggeri<sup>9</sup>, PP Piccaluga<sup>10</sup>, S Pileri<sup>10</sup>, E Tiacci<sup>11</sup>, B Falini<sup>11</sup>, B Pera-Gresely<sup>12</sup>, L Cerchietti<sup>12</sup>, J Iqbal<sup>13</sup>, WC Chan<sup>14</sup>, LD Shultz<sup>15</sup>, I Kwee<sup>5,16,17</sup>, R Piva<sup>1,18</sup>, I Wlodarska<sup>4</sup>, R Rabadan<sup>2</sup>, F Bertoni<sup>5,19</sup>, G Inghirami<sup>3,18,20</sup> and The European T-cell Lymphoma Study Group<sup>23</sup>



### **ALCL: ALK-negative**



1. Mason D, et al. In: Swerdlow SH, et al. World Health Organization (WHO) Classification of Tumours of Haematopoietic and Lymphoid Tissues, Fourth edition. Lyon, France: IARC Press; 2008, pp. 317; 2. Jaobsen E, The Oncologist 2006, 11:831-840.



1250

#### **CD30-positive peripheral T-cell lymphomas share molecular and phenotypic features**

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FoxP1 Syk EP573Y pSTAT3 C/EBPB Cyclin D3 IMP3 JunB MUM1/IRF4 Syk C-20 GATA1 TCR<sub>BF1</sub> ICOS CD69 CD52 NFATc2 **ZAP-70** CD3 Itk Lck Fyn MAL CD30+ ALK CD30-ALK+ ALCL PTCL, NOS PTCL, NOS ALCL

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haematologica | 2013; 98(8)





#### de Leval L and Gaulard P. Haematologica 2013; 95:1627-30.

#### Intracellular TCR-signaling Pathway Novel Markers for Lymphoma Diagnosis and Potential Therapeutic Targets

(Am J Surg Pathol 2014;38:1349-1359)

Claudio Agostinelli, MD, PhD,\* Hasan Rizvi, MD,† Jennifer Paterson, MS,† Vishvesh Shende, PhD,† Ayse U. Akarca, MS,† Elena Agostini, MD,\* Fabio Fuligni, MS,\* Simona Righi, MS,\* Sebastiano Spagnolo, BS,\* Pier Paolo Piccaluga, MD, PhD,\* Edward A. Clark, PhD,‡ Stefano A. Pileri, MD, PhD,\* and Teresa Marafioti, MD, PhD†





#### Gene expression analysis of peripheral T cell lymphoma, unspecified, reveals distinct profiles and new potential therapeutic targets

Pier Paolo Piccaluga,<sup>1,2</sup> Claudio Agostinelli,<sup>1</sup> Andrea Califano,<sup>3</sup> Maura Rossi,<sup>1</sup> Katia Basso,<sup>2</sup> Simonetta Zupo,<sup>4</sup> Philip Went,<sup>1,5</sup> Ulf Klein,<sup>2</sup> Pier Luigi Zinzani,<sup>1</sup> Michele Baccarani,<sup>2</sup> Riccardo Dalla Favera,<sup>2,6</sup> and Stefano A. Pileri<sup>1</sup>

JCI, 117:823-34, 2007

#### Gene expression profiling uncovers molecular classifiers for the recognition of

Anaplastic Large Cell Lymphoma within Peripheral T-cell neoplasms

JCO, 2010; 28:1583-90.

Roberto Piva<sup>1,2</sup>, Luca Agnelli<sup>3\*</sup>, Elisa Pellegrino<sup>1\*</sup>, Katia Todoerti<sup>3</sup>, Valentina Grosso<sup>1</sup>, Ilaria Tamagno<sup>1</sup>, Alessandro Fornari<sup>1</sup>, Barbara Martinoglio<sup>4</sup>, Enzo Medico<sup>4</sup>, Alberto Zamò<sup>5</sup>, Fabio Facchetti<sup>6</sup>, Maurilio Ponzoni<sup>7</sup>, Eva Geissinger<sup>8</sup>, Andreas Rosenwald<sup>8</sup>, Hans Konrad Müller-Hermelink<sup>8</sup>, Cristiane De Wolf-Peeters<sup>9</sup>, Pier Paolo Piccaluga<sup>10</sup>, Stefano Pileri<sup>10</sup>, Antonino Neri<sup>3</sup>, Giorgio Inghirami<sup>1,2</sup>

#### Identification of a three-gene model as a powerful diagnostic tool for the recognition of ALK negative ALCL Blood, 2012;120:1274-81.

Luca Agnelli, Elisabetta Mereu, Elisa Pellegrino, Tania Limongi, Ivo Kwee, Elisa Bergaggio, Maurilio Ponzoni, Alberto Zamò, Javeed Iqbal, Pier Paolo Piccaluga, Antonino Neri, John C. Chan, Stefano Pileri, Francesco Bertoni, Giorgio Inghirami and Roberto Piva





### Gene expression signatures delineate biologic and prognostic subgroups in peripheral T-cell lymphoma

Javeed Iqbal, George Wright, Chao Wang, Andreas Rosenwald, Randy D. Gascoyne, Dennis D. Weisenburger, Timothy C. Greiner, Lynette Smith, Shuangping Guo, Ryan A. Wilcox, Bin Tean Teh, Soon Thye Lim, Soon Yong Tan, Lisa M. Rimsza, Elaine S. Jaffe, Elias Campo, Antonio Martinez, Jan Delabie, Rita M. Braziel, James R. Cook, Raymond R. Tubbs, German Ott, Eva Geissinger, Philippe Gaulard, Pier Paolo Piccaluga, Stefano A. Pileri, Wing Y. Au, Shigeo Nakamura, Masao Seto, Francoise Berger, Laurence de Leval, Joseph M. Connors, James Armitage, Julie Vose, Wing C. Chan and Louis M. Staudt

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(A)	ALC	Ľ		EN	KTL		
AITL	ALK-	ALK+	ATTL	NK	γδΤ	PTCL-NOS	
			11	No.			





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JOURNAL OF CLINICAL ON	ICOLOGY	ORIGINAL REPORT		
Molecular Profiling Improves Classification and				
Prognostication of Nodal Peripheral T-Cell Lymphomas:				
Results of a Phase III Diagnostic Accuracy Study				
Pier Paolo Piccaluga, Fabio Fuligni, Antonio De Leo, Clara Bertuzzi, Maura Rossi, Francesco Bacci, Elena Sabattini, Claudio Agostinelli, Anna Gazzola, Maria Antonella Laginestra, Claudia Mannu, Maria Rosaria Sapienza, Sylvia Hartmann, Martin L. Hansmann, Roberto Piva, Javeed Iqbal, John C. Chan, Denis Weisenburger, Julie M. Vose, Monica Bellei, Massimo Federico, Giorgio Inghirami, Pier Luigi Zinzani, and Stefano A. Pileri				



### FFPE GEP effective in discriminating PTCL subtypes







### CD30-positive PTCL/NOS cases were classified as PTCL/NOS

- CD30-positive
  PTCL/NOS cases
  - No criteria for ALKnegative ALCL diagnosis
  - 16 cases
- Molecular classifier -  $- 16/16 \rightarrow \text{PTCL/NOS}$

#### No ALCL morphology; CD30 >75%



### ALK-ALCL vs. CD30+ PTCL/NOS

Blood, 15 June 2008, Vol. 111, No. 12, pp. 5496-5504.

ALK<sup>-</sup> anaplastic large-cell lymphoma is clinically and immunophenotypically different from both ALK + ALCL and peripheral T-cell lymphoma, not otherwise specified: report from the International Peripheral T-Cell Lymphoma Project

Kerry J. Savage<sup>1</sup>, Nancy Lee Harris<sup>2</sup>, Julie M. Vose<sup>3</sup>, Fred Ullrich<sup>4</sup> , Elaine S. Jaffe<sup>5</sup>, Joseph M. Connors<sup>1</sup>, Lisa Rimsza<sup>6</sup>, Stefano A. Pileri<sup>7</sup> , Mukesh Chhanabhai<sup>8</sup>, Randy D. Gascoyne<sup>8</sup>, James O. Armitage<sup>3</sup> , Dennis D. Weisenburger, for the International Peripheral T-Cell Lymphoma Project<sup>9</sup>





#### microRNA expression profiling identifies molecular signatures associated with anaplastic large cell lymphoma

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#### **Key Points**

- Anaplastic large-cell lymphoma has a unique miRNA signature.
- The miR-17~92 is an important downstream effector of ALK oncogenic pathway.

Anaplastic large-cell lymphomas (ALCLs) encompass at least 2 systemic diseases distinguished by the presence or absence of anaplastic lymphoma kinase (ALK) expression. We performed genome-wide microRNA (miRNA) profiling on 33 ALK-positive (ALK[+]) ALCLs, 25 ALK-negative (ALK[-]) ALCLs, 9 angioimmunoblastic T-cell lymphomas, 11 peripheral T-cell lymphomas not otherwise specified (PTCLNOS), and normal T cells, and demonstrated that ALCLs express many of the miRNAs that are highly expressed in normal T cells with the prominent exception of miR-146a. Unsupervised hierarchical clustering demonstrated distinct clustering of ALCL, PTCL-NOS, and the AITL subtype of PTCL. Cases of ALK(+) ALCL and ALK(-) ALCL were interspersed in unsupervised

analysis, suggesting a close relationship at the molecular level. We identified an miRNA signature of 7 miRNAs (5 upregulated: miR-512-3p, miR-886-5p, miR-886-3p, miR-708, miR-135b; 2 downregulated: miR-146a, miR-155) significantly associated with ALK(+) ALCL cases. In addition, we derived an 11-miRNA signature (4 upregulated: miR-210, miR-197, miR-191, miR-512-3p; 7 downregulated: miR-451, miR-146a, miR-22, miR-455-3p, miR-455-5p, miR-143, miR-494) that differentiates ALK(-) ALCL from other PTCLs. Our in vitro studies identified a set of 32 miRNAs associated with ALK expression. Of these, the miR-17~92 cluster and its paralogues were also highly expressed in ALK(+) ALCL and may represent important downstream effectors of the ALK oncogenic pathway. (*Blood*. 2013;122(12):2083-2092)



Next-Generation Sequencing Identifies Deregulation of MicroRNAs Involved in Both Innate and Adaptive Immune Response in ALK+ ALCL PLOS ONE | DOI:10.1371/journal.pone.0117780 February 17, 2015

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miRNA	ALK- ALCL	ALK+ ALCL	fold change	padj
hsa-miR-196b	186	0	0,0006	2.48E-43
hsa-miR-155	44475	1205	0,0271	8.56E-35
hsa-miR-340	0	1538	3660,0000	8.56E-35
hsa-miR-146a	2525	29	0,0114	3.34E-27
hsa-miR-203	0	316	751,0000	1.19E-23
hsa-miR-135b	2	405	172,0000	1.07E-21
hsa-miR-424	1875	98	0,0525	3.22E-21
hsa-miR-135b*	0	119	562,0000	1.67E-18
hsa-miR-503	343	22	0,0645	1.14E-16
hsa-miR-424*	910	78	0,0856	7.64E-15
hsa-miR-182	672	17849	26,6000	3.79E-14
hsa-miR-183	51	1473	28,7000	3.79E-14
hsa-miR-542-3p	413	38	0,0912	1.40E-13





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#### ALK-negative anaplastic large cell lymphoma is a genetically heterogeneous disease with widely disparate clinical outcomes

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#### **Key Points**

- ALK-negative ALCLs have chromosomal rearrangements of *DUSP22* or *TP63* in 30% and 8% of cases, respectively.
- DUSP22-rearranged cases have favorable outcomes similar to ALK-positive ALCLs, whereas other genetic subtypes have inferior outcomes.



#### **DUSP22-IRF4 locus**



### **PRDM1/BLIMP1** is commonly inactivated in anaplastic large T-cell lymphoma

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#### **Key Points**

- The commonest lesions in anaplastic large cell lymphomas are losses at 17p13 and at 6q21, concomitant in up to onequarter of the cases.
- PRDM1 (BLIMP1) gene (6q21) is inactivated by multiple mechanisms and acts as a tumor suppressor gene in anaplastic large B-cell lymphoma.

Anaplastic large cell lymphoma (ALCL) is a mature T-cell lymphoma that can present as a systemic or primary cutaneous disease. Systemic ALCL represents 2% to 5% of adult lymphoma but up to 30% of all pediatric cases. Two subtypes of systemic ALCL are currently recognized on the basis of the presence of a translocation involving the anaplastic lymphoma kinase *ALK* gene. Despite considerable progress, several questions remain open regarding the pathogenesis of both ALCL subtypes. To investigate the molecular pathogenesis and to assess the relationship between the ALK<sup>+</sup> and ALK<sup>-</sup> ALCL subtypes, we performed a genome-wide DNA profiling using high-density, single nucleotide polymorphism arrays on a series of 64 cases and 7 cell lines. The commonest lesions were losses at 17p13 and at 6q21, encompassing the *TP53* and *PRDM1* genes, respectively. The latter gene, coding for BLIMP1, was inactivated by multiple mechanisms, more frequently, but not exclusively, in ALK<sup>-</sup>ALCL. In vitro and in vivo experiments showed that that *PRDM1* is a tumor suppressor gene in ALCL models, likely acting as an antiapoptotic agent. Losses of *TP53* and/or *PRDM1* were present in 52% of ALK<sup>-</sup>ALCL, and in 29% of all ALCL cases with a clinical implication. (*Blood.* 2013;122(15):2683-2693)



### 17p13 (TP53) – 6q21 (PRDM1)\*

#### Α

Lesion	freq	fn
ALK translocation	52%	
17p13.3-p12 loss	25%	
6q21 loss	19%	
1q gain	23%	
16q23.1-q23.2 loss	16%	
1p13.3-p12 loss	11%	
13q32.2-q33.3 loss	16%	
8q24.22 gain	17%	
10p11.23-p11.22 loss	14%	
1p36.33-p36.32 loss	13%	
7q gain	14%	
7p gain	11%	
12p13.33-p13.31 gain	13%	
2p25.3-p25.2 gain	11%	

100%

9%

3% 15%

3%

0%

9%

12%

6%

6%

12%

12%

9%

9%





# Gains of 9p24.1 (JAK) and 13q31.3 (MIR17HG)



#### Convergent Mutations and Kinase Fusions Lead to Oncogenic STAT3 Activation in Anaplastic Large Cell Lymphoma

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A systematic characterization of the genetic alterations driving ALCLs has not been performed. By integrating massive sequencing strategies, we provide a comprehensive characterization of driver genetic alterations (somatic point mutations, copy number alterations, and gene fusions) in ALK<sup>-</sup> ALCLs. We identified activating mutations of *JAK1* and/or *STAT3* genes in ~20% of 155 ALK<sup>-</sup> ALCLs and demonstrated that 38% of systemic ALK<sup>-</sup> ALCLs displayed double lesions. Recurrent chimeras combining a transcription factor (*NFkB2* or *NCOR2*) with a tyrosine kinase (*ROS1* or *TYK2*) were also discovered in WT JAK1/STAT3 ALK<sup>-</sup> ALCL. All these aberrations lead to the constitutive activation of the JAK/STAT3 pathway, which was proved oncogenic. Consistently, JAK/STAT3 pathway inhibition impaired cell growth in vitro and in vivo.







#### Clinical Advisory Committee Meeting WHO Classification Update Chicago March 31 – April 1, 2014





### In the updated WHO Classification

### ALK<sup>-</sup> ALCL

### **Distinct entity!**

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#### Intralymphatic Cutaneous Anaplastic Large Cell Lymphoma/Lymphomatoid Papulosis Expanding the Spectrum of CD30-positive Lymphoproliferative Disorders

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# Thank you very much for your attention!